

22nd Annual Symposium
Clinical & Pharmaceutical Solutions through Analysis

Microsampling & Patient Centric Sampling

A journey through what it is and how you can incorporate it into your workflows

Monday 28th October 2019, 12:00-16:00 University Grille

Clinical & Pharmaceutical Solutions through Analysis October 28-31, 2019 Langhorne, PA

Short Course Outline

12:00	Lunch, Welcome & Introductions Neil Spooner & Joe Siple
12:30	Introduction to Microsampling Presentation - Neil Spooner
13:00	Considerations for drug bioanalytical assay method development & validation, and clinical implementation strategies Presentation & Discussion - Tim Olah & Enaksha R Wickremsinhe
14:00	Break
14:15	Considerations for Clinical Operations Presentation & Discussion - Melanie Anderson
15:15	Future Directions Discussion - Kevin Bateman & Neil Spooner
15:45	Wrap-up & next steps
16:00	End



Introduction to Microsampling

Neil Spooner PhD, CChem, FRSC (neil@spoonerbioanalytical.co.uk)

Founder & Director - Spooner Bioanalytical Solutions Ltd, UK
Senior Visiting Research Fellow - School of Life & Medical Sciences, University of Hertfordshire, UK
Senior Editor — Bioanalysis Journal

Microsampling: considerations for its use in pharmaceutical drug discovery and development

Neil Spooner*,¹, Kenneth D Anderson², Joe Siple³, Enaksha R Wickremsinhe⁴, Yang Xu² & Mike Lee⁵

There is growing interest in the implementation of microsampling approaches for the quantitation of circulating concentrations of analytes in biological samples derived from nonclinical and clinical studies involved in drug development. This interest is partly due to the ethical advantages of taking smaller blood volumes, particularly for studies in rodents, children and the critically ill. In addition, these technologies facilitate sampling to be performed in previously intractable locations and occasions. Further, they enable the collection of samples for additional purposes (extra time points, biomarkers, sampling during a clinical event, etc). This article gives a comprehensive insight to the utilization of these approaches in drug discovery and development, and provides recommendations for best practice for nonclinical, clinical and bioanalytical aspects.

First draft submitted: 26 February 2019; Accepted for publication: 30 April 2019; Published online: 20 June 2019

¹Spooner Bioanalytical Solutions Ltd, Hertford, UK & School of Life & Medical Sciences University of Hertfordshire, UK

²Merck & Co. Inc., Department of Pharmacokinetics, Pharmacodynamics & Drug Metabolism, West Point, PA 19486, USA

³New Objective Inc, 2 Constitution Way, Woburn, MA 01801, USA

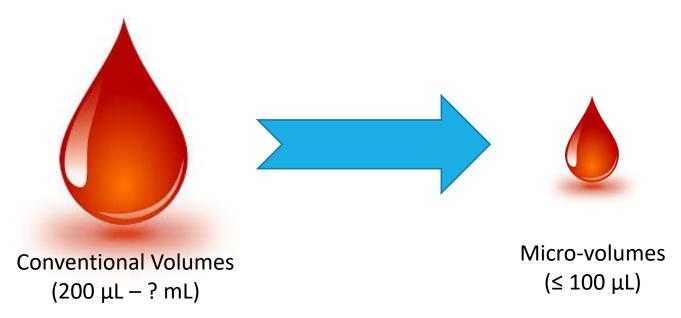
⁴Lilly Research Laboratories, Eli Lilly & Company, Indianapolis, IN, USA

Milestone Development Services, PO Box 178, Newtown, PA 18940-0178, USA

^{*}Author for correspondence: neil@spoonerbioanalytical.co.uk



What is microsampling?



Technologies for collecting & analysing smaller blood & plasma / serum volumes for the accurate determination of circulating concentrations of therapeutic drugs, metabolites & biomarkers in non-clinical & clinical studies

Spooner et al (2019) Bioanalysis 11(10) 1015–1038

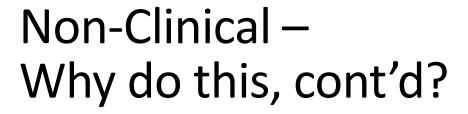
Non-Clinical – Why do this?



Ethical - 3Rs

- Reduction in rodent animal numbers
 - Elimination of TK satellites reduces number of animals by 30-40%
 - Effects primarily on reticulocytes; no affect in overt toxicity assessment,
 e.g., hepatotoxicity, renal toxicity*
 - Serial TK & PK sampling in mice
 - Discovery PK/PD, mouse TK & PK/PD & juvenile studies
- Refinement of bleeding technique
 - Reduction, or elimination of rodent warming
 - Sampling from more convenient / less disruptive location

^{*}Powles-Glover et al (2014) Reg. Toxicol. Pharmacol. 68, 325-331





Improved data quality

- Exposure data in main study animals, rather than additional satellites
- Direct correlation of exposure with PD and toxicological outcomes

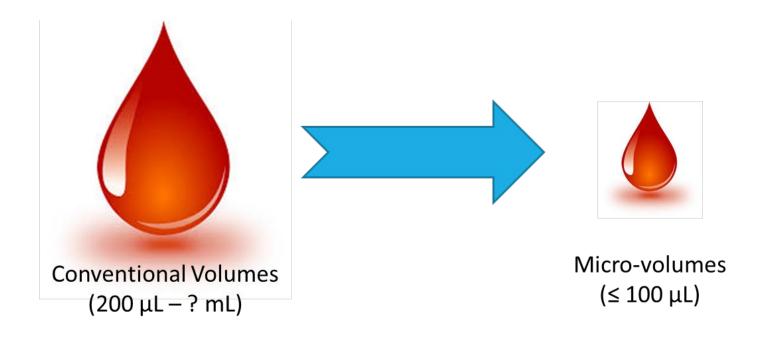
Enables samples to be taken for other purposes

 Additional PK/TK timepoints, biomarkers, metabolites, Clin. Path. determinations, etc.

Cost

- Reduced animal numbers, housing, drug substance
 -but, consumable costs are higher

See https://nc3rs.org.uk/microsampling



However, this course is **NOT** simply about microsampling...



....But it is about moving beyond conventional clinical blood sampling



It's about collecting.....

...the appropriate sample...

...in a location that is most convenient for the patient...

...that provides high quality information

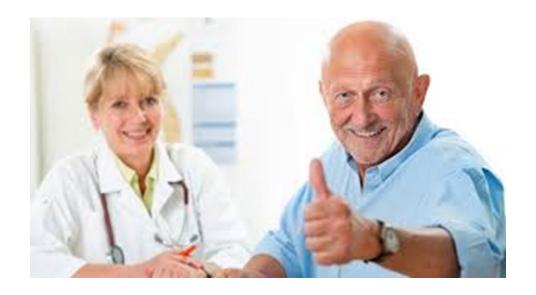


This may be blood sample volumes of 10 μ L, or it may be 250 μ L





With the **PATIENT** at the centre of our considerations



Patient Centric Sampling





Quality

Obtaining a high quality blood / plasma / serum sample for accurate quantitative determination of drugs, drug metabolites & endogenous molecules

Patient

Minimising the impact on the human patient / consumer

- Optimising blood volume sampled
- Minimising pain
- Facilitating convenience

New Data

Generating concentration data in situations that are currently difficult, or impossible to work with



Dried Blood Spots

Established for neonatal screening for 50+ years

Delivers all the benefits outlined

PLUS - Simpler process

- Removes need for centrifugation or sub-aliquots
- Dry ice and freezers not required
 - BIG cost savings on sample shipments

Barfield *et al* (2008) *J. Chrom. B* **870**, 32-37; Spooner *et al* (2009) *Anal. Chem.* **81**, 1557-1563; Spooner *et al* (2010) *Bioanalysis* **2(8)** 1515-1522; Pandya *et al* (2011) *Bioanalysis* **3(7)** 779-786; Stokes *et al* (2011) *Lab. Animals* **45**, 109-113;



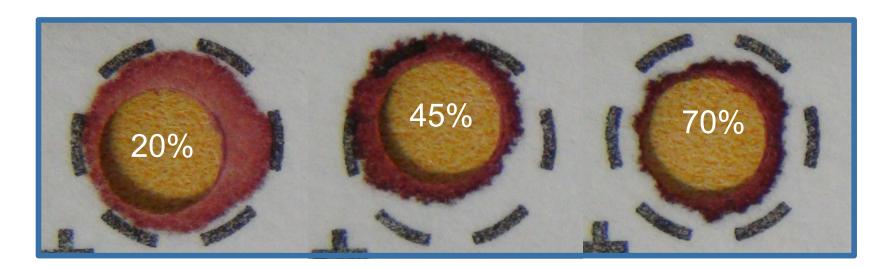




However!!!

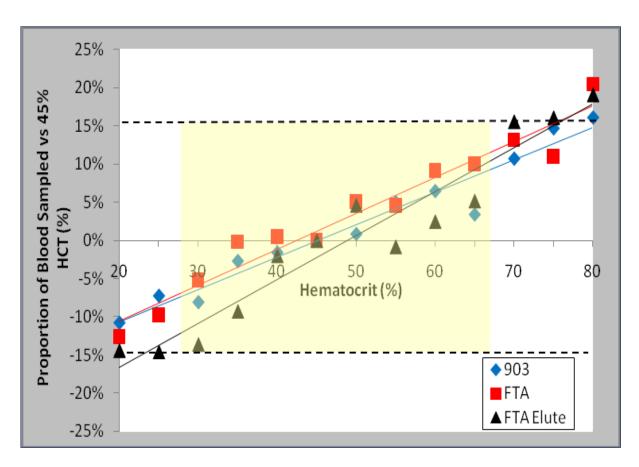


Blood hematocrit affects the size of the derived blood spot



Leading to a bias in the quantitative data!





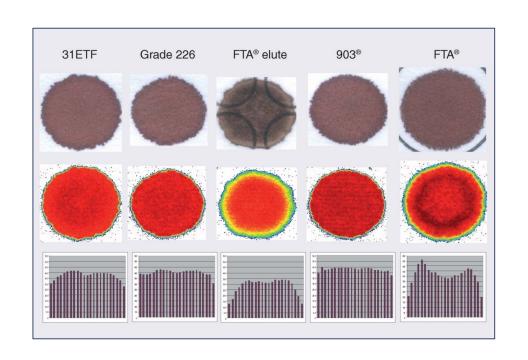
Fixed diameter disc collected from spot with varying HCT

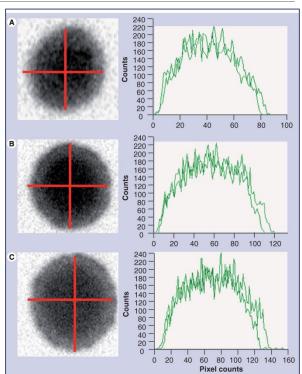
 All data normalized to 45% HCT

Denniff & Spooner (2010) Bioanalysis **2(8)** 1385-1395



Spot homogeneity





Example radio histograms of the (A) 15-, (B) 30- and (C) 45- μ l blood spots spiked with 14 C radiolabeled UK-414495

Ren et al, (2010) Bioanalysis 2(8) 1469-1475; Clark et al (2010) Bioanalysis 2(8) 1477-1488



Resulting in.....

Regulators (FDA & EMA) required collection & analysis of both wet and dry samples and demonstration of concordance in healthy volunteers and patient groups



Denniff & Spooner (2010) *Bioanalysis* **2(8)** 1385-1395; O'Mara *et al* (2011) *Bioanalysis* **3(20)** 2335-2347; de Vries *et al* (2013) *Bioanalysis* **5(17)** 2147-2160; Cobb *et al* (2013) *Bioanalysis* **5(17)** 2161-2169; Evans *et al* (2015) *AAPS J.* **17(2)** 292-300; Kothare *et al* (2016) *AAPS J.* **18(2)** 519–527

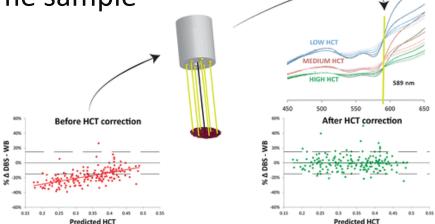


Ways forward for DBS sampling

 Closely match HCT of analytical calibrants & QC's to that of the clinical study samples

Normalise data to another readily measured component

of the same sample



Collect accurate sample volume & analyse entire sample

Capiau et al (2018) Anal. Chem. 90(3) 1795-1804; Velghe et al (2019) J. Pharm. Biomed. Anal. 163, 188-196

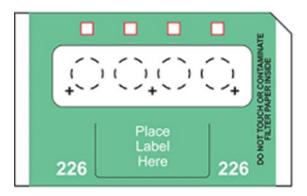




For quantitative analysis

- Technologies required that overcome the issues associated with
 - Blood hematocrit
 - Sample homogeneity
- Whilst delivering the benefits
 - Collecting smaller blood volumes (where appropriate)
 - Facilitating self/assisted sampling
 - Delivering cost savings through home sampling & room temperature sample shipments
 - Integrating with systems for sample shipping, tracking & analysis







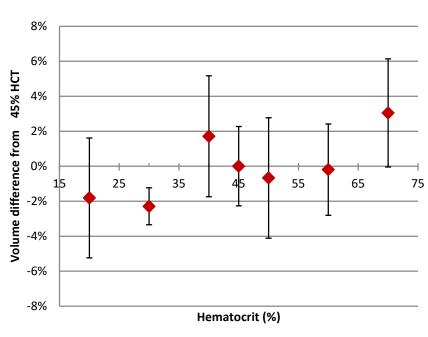




HCT independent volumetric sampling performance







- Human blood at different HCTs was spiked with ¹⁴C caffeine
- Tip oxidised to CO₂

Denniff & Spooner (2014) *Anal. Chem.* **86**, 8489-8495, Denniff *et al* (2015) *J. Pharm. Biomed. Anal.* **108**, 61-69, Spooner *et al* (2015) *Bioanalysis* **7(6)** 653-659

This is **NOT JUST** about collecting the samples and data we do today!



Pediatrics

Critically ill

Remote areas

Additional data

- PK
- Biomarkers
- Compliance
- Therapeutic drug monitoring
- Medical event migraine
- Longitudinal



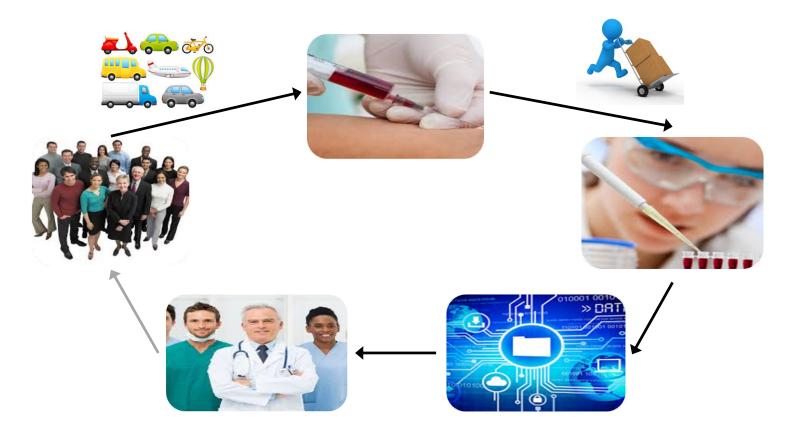
Improved clinical trial recruitment & retention

Patient / Consumer driven healthcare



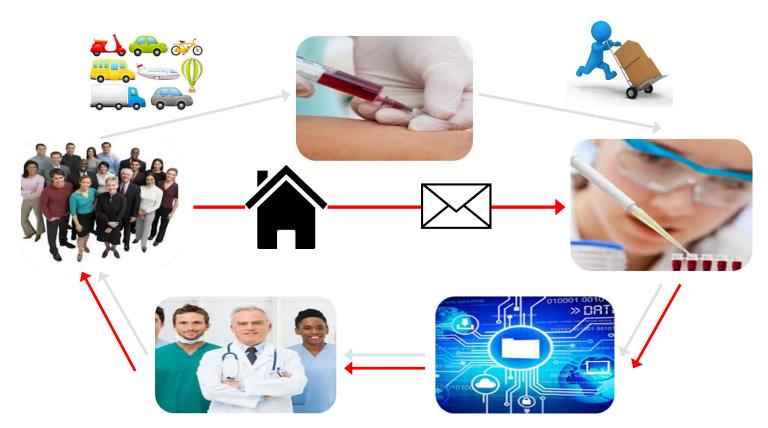








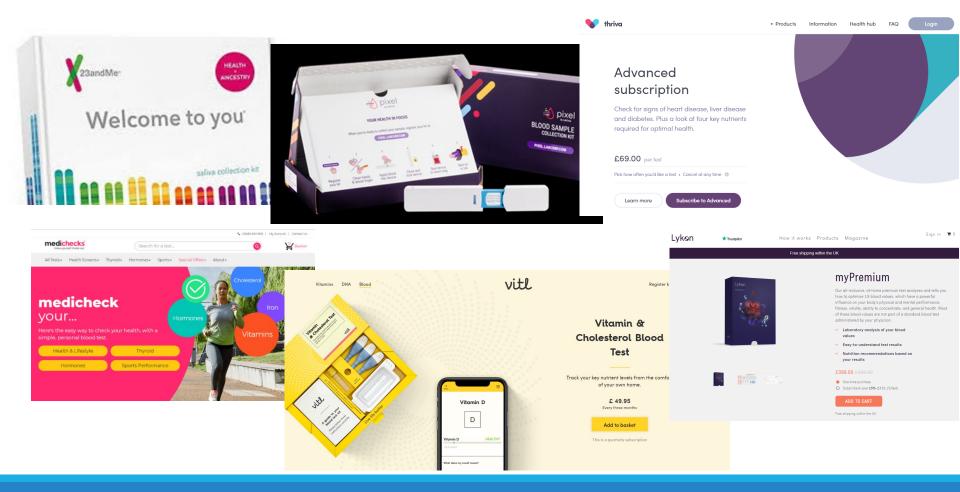
Future?



More often, for more analytes.....

Is the future already here? Consumer-led healthcare







Potential challenges.....

Is the sample representative of the whole?

Non-conventional sample format

- Sample transfer
- Storage
- Analysis
- Automation

Bridging to existing data

- Blood / Plasma
- Venous / Capillary
- Wet / Dry

Additional assay validation steps

Data acceptability

Regulators

Reluctance to change!!!





Regulatory Landscape

Non-Clinical

- ICH Q&A on Microsampling as part of ICH S3A Guideline (Nov 2017)
- Also See
 - Beharry (2010) Bioanalysis 2(8), 1363–1364
 - Viswanathan (2012) Bioanalysis 4(12), 1417–1419

Clinical

- FDA guidance provided in latest <u>BMV document</u> (May 2018)
- Draft ICH M10 Guidance on BMV (Feb 2019)
- Also see
 - Evans, et al (2014) The AAPS Journal 17(2), 292-300
 - Kothare, et al (2016) The AAPS Journal 18(2), 519–527

Further details on BMV will be given later in this course



Change is difficult!



It depends on how you look at it!









Patient Centric Sampling Interest Group



>90 members

>50 different organisations

 CROs, biotech, pharma, device innovators, instrument vendors, consumable vendors, consultancies, etc

Collaborate in non-competitive areas of interest

- Standardisation
 - Working with CLSI to build an industry standard
- Broad acceptance of patient centric sampling technologies and pathways to their implementation
 - Public, Scientific community, Medical community, Legislators, Payers, Regulators, Media, etc......
 - Building external facing website

Other Forums for working together

















Conclusions

New blood sampling & analytical technologies are emerging

Enables us to put the patient first & facilitates human wellbeing

Enables us to collect data that has previously been difficult / impossible to obtain

It's about so much more than the devices

Working across boundaries will enable the change to happen

Change will not be easy & there will be surprises



Clinical & Pharmaceutical Solutions through Analysis October 28-31, 2019 Langhorne, PA

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Microsampling Workshop

Considerations for bioanalytical assay method development/validation and clinical implementation strategies for drug candidates

Enaksha Wickremsinhe & Tim Olah

Selecting a Microsampling technique

- Multiple techniques: enabling collection of reduced blood volumes
- Stage of assay implementation: Discovery or Development
- <u>Discovery</u>: minimal validation, fit for purpose
- <u>Development</u>: must meet BMV guidance (EMA 2011, FDA 2018, ICH M10, etc)
- What is different when compared to routine bioanalytical methods
 - Dried sample (blood or plasma) vs wet sample (blood or plasma)
 - Sample volume: capability to handle small samples/volumes
 - Collection device/format: selection, ease of use, cost
 - **Source** of blood: IV draw, finger-stick, subcutaneous



Bioanalytical challenges for microsampling

- Preparation of Standard Curves and QCs
- Assay sensitivity: can you achieve the required LLOQ?
- Additional validation experiments: depends on technique employed
- Account for stability during collection/transit/storage
 - temperature, humidity, drying time, shipping conditions, etc.
- Addition of Internal Standard: in extraction solvent or on pre-dried device?
- More time and effort required in BioAnalytical lab
 - Samples are <u>not</u> in 96-well format, AUTOMATION not currently possible
- Sample storage and related logistics: physical change or analyte degradation
- Overall BioAnalytical cost higher than current practices?

Other Bioanalytical challenges

which may require unique experiments or assessments

- Cost to perform cross validation
- Cost of "novel" devices needed for method dev and validation
- Impact of mismatched data sets (poor correlation?)
- Understanding assay efficiencies, operational errors?
- Impact of shipping, storage, and handling temperatures
- Assess homogeneity of samples (especially DBS)
- Multiple "aliquots" of microsamples



Considerations for Clinical Implementation

- In Vitro data: B:P ratio and Fu (protein binding) should not be conc dependent. No Hct effect over clinically relevant range
- 2. <u>BioA feasibility</u>: LLOQ, Stability, Hct, Homogeneity, etc.
- 3. Establishing concordance (Bridging): relationship between microsampling conc data and traditional sample (plasma/serum) concentrations data (such that PK conclusions can be drawn across studies)

Is Microsampling data = IV blood/plasma/serum data?

Typically, the biological matrix is either liquid plasma (SM) or serum (LBA)

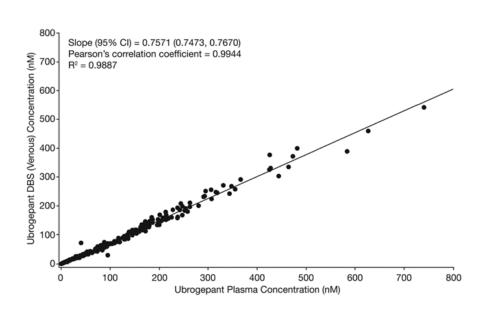
- Microsampling introduces additional matrices
 - Plasma collected in capillaries
 - Liquid blood
 - Blood diluted in water
 - Dried blood (DBS, VAMS, etc.)
 - Dried plasma
 - Other dried matrices: urine, CSF, etc.
- Also provides alternative sampling sites (compared to IV draw)
 - Finger stick
 - Sub-cutaneous (Tasso, TAP etc)
 - Arterial blood: via umbilical catheter in neonates.

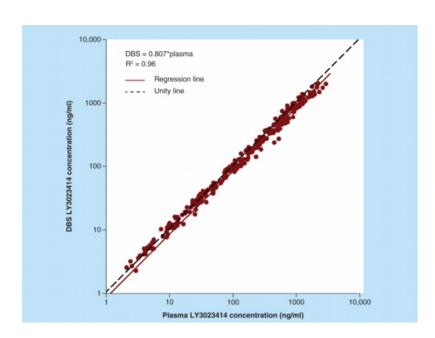


Establish concordance (cross validation) with traditional method (plasma/serum)

- 2018 FDA BMV provides guidance on demonstrating correlation between the microsampling method and traditional method
 - Wet vs Dry
 - Plasma vs Blood
 - Venous blood/plasma vs finger stick blood/plasma
- Use incurred samples (n > 20)?
- Blood from Healthy volunteers? Patients?
- Acceptance criteria?
- Seek feedback from Regulatory agency (FDA).







Li, Bateman, Kothare, et al, 2018. J Clin Pharmacol, Vol. 58, No. 3, 294-303

Wickremsinhe et al., Bioanalysis (2018) 10(5), 341-356



How do you decide: to microsample or not!

It's not a panacea: Just another tool in the BioA tool box.

Make sure it's the right one for the right study

Needs input from the following:

- Medical
- Clinical operations
- PK
- ADME/DMPK
- BioAnalytical





Discussion Questions

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Break

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Clinical Operations

How to implement patient centric sampling in clinical trials



What is the need? What question are we answering? Can micro sampling meet this need?

- Program/Institutional Needs
 - Matrix requirements
 - At home sampling benefits
 - Specific indications pediatrics, migraine, cancer, therapeutic drug monitoring

The technology is disruptive to existing workflows across the organization – the need must be great



Logistical

- Training Clinical site and Patient this can involve several clinical sites all over the world and require language translation.
- Sample Integrity/Quality how do we ensure the right person gets sampled at the right time in the correct way?
- Environmental Exposure in the wild
- Technology access for use in remote/underserved geographies if using an eDiary/App based data collection approach is used.
- Supply scaling up manufacturing for device availability, lot-to-lot variability.
 - How do you track various lots across large clinical studies, make standards and QCs with matching lots, etc.?
- Shipping requirements within a country and country to country?



Logistical

- How do we reliably collect a time stamp and how will the data flow?
 - No preassigned barcodes for sample collection, new process needs to be developed and proven to work.
 - How do you match the concentration values with the date and time of collection?
 - What about time zones?
- Patient population may not be appropriate for use of invasive technologies (i.e. shipping HIV infected blood).
- Patient compliance and sample collection reliability, at home sampling needs to be as simple and straightforward as possible.





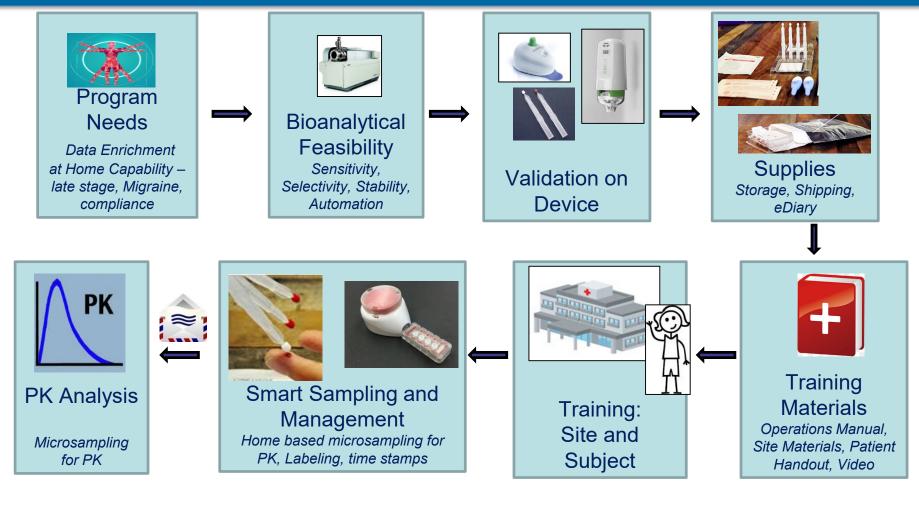


- Records time and temperature every 10 min for 2 weeks
- Starts when button is pressed
- wireless
 communication with
 smartphone or
 smartbox (to be
 design in partnership
 with Merck)

Business/Regulatory Related

- "If it can't be used at 100% of sites, it can't be used at all" attitude.
- Increases the cost of conducting the trial.
- Requires bridging from liquid plasma to dried blood.
- Increases the complexity of the protocol for the trial and this will impact enrollment.
- No definitive data that shows return on investment for Patient-Centric Sampling.
- How are devices treated and what regulatory approval is needed in each country?
- How do we engage with regulators to minimize issues during filing?
- How do you show the sample is from the person enrolled in the trial?
- Can you define inclusion/exclusion criteria using adherence data from at home sampling? What about intent to treat criteria?
 - Potential for un-blinding when using these approaches











Supplies/Kits

Patient Supply Kit

User guide

Alcohol wipe

Lancet

Sterile gauze pad

Band-Aid®

VAMS tip clamshell

Foil Envelope

Desiccant

Mailing Envelope



Day 13: At Home What you need to do today: Charge iPhone and if

 Refer to the CleverCap User Guide on how to remove your study drug from the bottle
 1: Take your study drug between 6:00 a

Step 2: Turn on the iPad, open the Proteus application.

and time you took the study drug

- The study staff will take extra blood samples from you today.
 The clinic site staff will collect samples.
- of blood from your arm at the same time you collect blood for the dried blood spot cards
- Charge iPhone and iPad and check patch status
- Self-Collected Fingerstick Dried Blood Spot Samples

 • Refer to CleverCap User
- Refer to CleverCap User Guide on how to remove yo study drug from the bottle



Sites 2: Takes your shady drug between 6:00 am and 10:00 am as directed by the shady staff.

Sites 3: Scan the bar code of the direct blood spot sample collection card on the e-diary (DBS Guide Step 7).

Sites 4: Record the date and time of blood sample collection in the corresponding fields of the e-diary.

Step 5: Set the card to dry (DBS Guide Step 8)

Step 6: Store the dried blood spot card (DBS Guide Step 9)

(DBS Guide Step 1-9)
Step 8: Two (2) hours after you take your drug collect your blood sample when reminder alarm goes

off (DBS Guide Step 1-9) Step 9: Four (4) hours after you take your drug -

Don't forget to bring all devices and envelopes containing the Dried Blood Spot Cards Checklist

iPad and patch (still wearing on your skin)
 iPhone (e-diary)
 CleverCap pill bottle and 2net Hub
 All envelopes containing DBS cards (Day 5, 8, and 11)

The clinic site staff will provide instructions fo the second part of the MK-0431-841 study

Collection Kits



Lancet

Alcohol wipe



WITCHES THE PARTY OF THE PARTY

VAMS Tip Clamshell



Foil Envelope and Desiccant



Band-Aid®

Sterile gauze pad

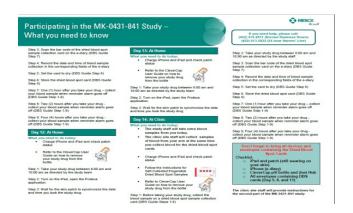






Training Materials

- Brochures for Patients
- Brochures for Clinical Staff
- Study Operations Manual
- Videos
- Investigator Meeting Presentations



6.2. Remove device from sterile pouch by pulling the two layers apart to open



- 6.3. Ensure that the device is intact (the transparent protective cover of the Tasso device and the adhesive backing are in place).
- 6.4. Reveal the application site this is located on the upper arm, 3 finger width-distance from the top of the shoulder, as shown in the diagram below:



6.5. Swab application site with provided ethanol swab. Allow site to air dry



6.7. Place the Tasso device on the swabbed application site with the collection pod pointed down.

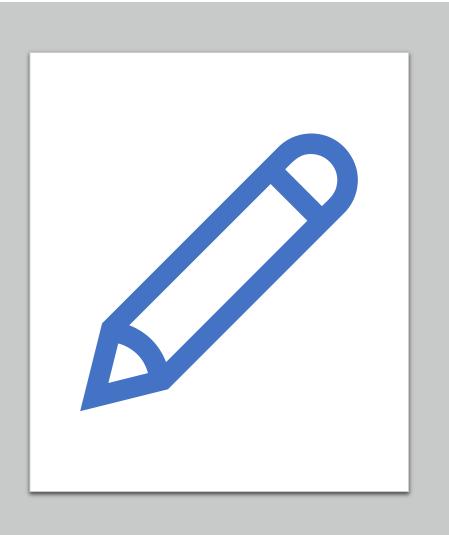


6.8. Remove the transparent Tasso device protective cover, revealing the activation button.



6.9. Press firmly and evenly with your thumb on the center of the Tasso button until you hear an audible click. Release your thumb and start the timer.





Future Directions

Kevin Bateman & Neil Spooner

What do we need to do in order to progress?

Technology

- Sampler features
 - Ease of use end user vs analytical scientist
 - Sample volume
 - Sample format
 - Wet, dry, plasma, serum, blood
 - Single vs replicates
 - Sample processing at point of collection
 - Time, date & location
 - Traceability
 - Recycle / re-use
- Analytical workflows
 - Centralised vs decentralised
 - Skilled analysts vs push button
 - Compatibility / integration with analytical methods
- Standardisation

What do we need to do in order to progress?

- Implementation
 - Organisational change
 - Within group
 - Between groups
 - Regulatory acceptance
 - Acceptance by society
 - Training
 - Patients
 - Clinicians
 - Analytical Scientists

Where else can we use these approaches?

- Large molecule bioanalysis
 - LBA and/or LC-MS approaches
- Vaccine research
 - Epidemiological studies
 - Track response/protection over time
- Non-drug analytes
 - Dynamics of disease signatures (RNA)
 - Longitudinal studies of human health

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Wrap-up and Next Steps

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